

SECONDARY METABOLITES BY
CHEMICAL SCREENING. PART 19[†]

SM 196 A AND B, NOVEL BIOLOGICALLY
ACTIVE ANGUICYCLINONES
FROM *Streptomyces* sp.

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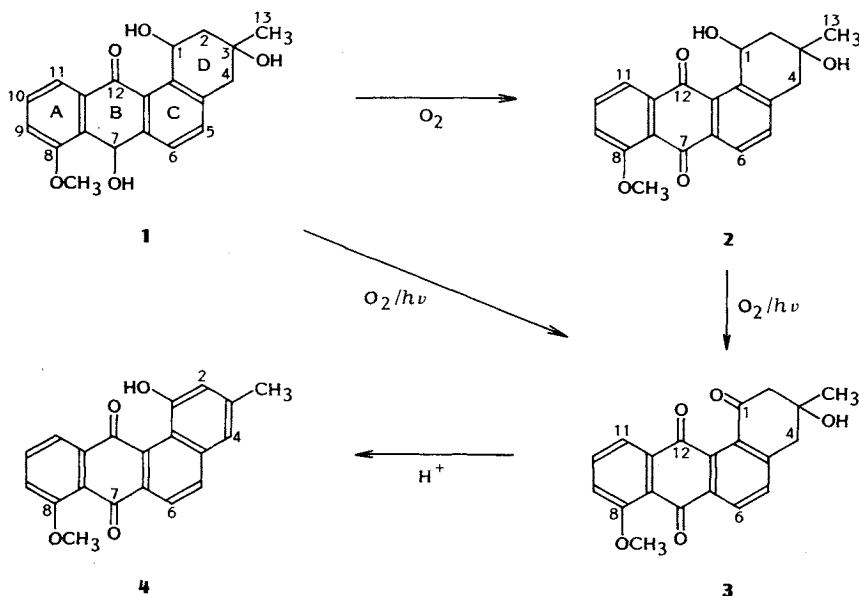
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Application of a method of chemical screening²⁾, which based on the prior work of ZÄHNER *et al.*³⁾, gave rise to the detection, isolation and structure elucidation of two new angucyclinones⁴⁾ exhibiting biological activity. In addition, the known compounds 6-deoxy-8-*O*-methylrabelomycin (3)^{5,6)} and X-14881 E (4)⁷⁾ were detected in the same culture broth.

The producing organism *Streptomyces* sp. (DSM 4769) was isolated from a soil sample collected near Ajanta (India) and exhibits tightly spiraled, red spores with a smooth surface. Melanin production is positive. Cultivation was carried out in a seed medium (meat flour 2%, malt extract 10%, CaCO₃ 1%, pH adjusted to 7.2 prior to sterilization) inoculated from an agar slant culture and incubated for 48 hours at 27°C. The resulted seed medium (5%) was used to inoculate the production medium (glycerol 3%, casein peptone 0.2%, K₂HPO₄ 0.1%, NaCl 0.1%, MgSO₄·7H₂O 0.05% and 5 ml of a mineral solution containing CaCl₂·2H₂O 3 g, FeC₆H₅O₇ 1 g, MnSO₄ 0.2 g, ZnCl₂ 0.1 g, CuSO₄·5H₂O 0.025 g, Na₂B₄O₇·10H₂O 0.02 g, Na₂MoO₄·2H₂O 0.01 g, and CoCl₂ 4 mg per liter water). Production was carried out in 300-ml Erlenmeyer flasks containing 100 ml of production medium (200 rpm on a rotary shaker, 30°C) or in 10-liter fermenters (250 rpm, aeration: 4 liters/minute) for 98 hours.

We prepared extracts from the culture filtrate following a procedure described elsewhere²⁾. Surprisingly, without the use of any staining reagent a colorless spot on the TLC plate (SM 196 A (1): R_f 0.25, CHCl₃-MeOH (15:1); Silica gel 60F₂₅₄ on glass, Merck, HPTLC-Fertigplatten) immediately turned to brilliant red after exposing the TLC plate

Scheme 1.



[†] See ref 1.

to UV light (366 nm). In addition, two minor yellow spots (SM 196 B (2); Rf 0.40; 3: Rf 0.33) changed their color to orange. Development of the TLC plate in a second dimension using the same solvent system showed 1 to be converted into 2, 3, and 4, whereas 2 was transformed to 3 and 4, and 3 gave 4 possibly by H⁺-catalysis.

The culture filtrate (8 liters) of a 10-liter fermenter was extracted three times with 5 liters CH₂Cl₂. The organic layer was evaporated to dryness and the remaining oily residue (4 g) was chromatographed on a silica gel column (wrapped with aluminum foil to prevent photo-conversion; 30 × 3 cm; CHCl₃-MeOH (30:1)) to obtain the four detected metabolites: colorless crystalline SM 196 A (1, 33 mg), yellow amorphous SM 196 B (2, 22 mg), orange amorphous 3 (21 mg), and red 4 (12 mg, Rf 1.0). Due to the chemical instability of the compounds this purification turned out to be difficult, especially in the case of SM 196 B (2). The new angucyclinones 1 and 2 are soluble in MeOH, acetone, DMSO, dioxane, and CHCl₃ and are insoluble in H₂O or *n*-hexane.

The structures of 1 and 2 were ascertained by ¹H, ¹³C and 2D NMR techniques. Some of the relevant ¹H and ¹³C NMR data are summarized in Tables 1 and 2. The compounds of the SM 196 complex exhibit a benz[*a*]anthracene carbon skeleton and therefore belong to the group of angucyclinones⁸. Based on these NMR data, compound 3 was proved to be identical with 6-deoxy-8-*O*-methylrabelomycin^{5,6} and 4 with X-14881 E⁷. The colorless

metabolite SM 196 A (1) possesses a dihydroquinone moiety in ring B and a secondary hydroxy group at C-1. FAB, CI (NH₃), and EI (70 eV) mass spectra of SM 196 A (1) showed peaks at *m/z* 338 (M⁺ - H₂) and 322 (M⁺ - H₂O).

Table 1. ¹³C NMR data of SM 196 A (1), B (2), and X-14881 E (4).

C-Atom	SM 196 A (1)	SM 196 B (2)	X-14881 E (4)
1	65.73	65.28	155.10
2	44.14	43.97	119.71
3	69.05	69.18	141.14
4	45.17	45.40	121.11
4a	136.43	142.15	130.67
5	135.70	136.05	137.53
6	128.81	127.27	122.76
6a	143.35	136.05	136.74 ^a
7	63.34	182.37	182.12
7a	129.36	120.60	119.15 ^b
8	156.81	159.71	159.57
9	114.78	117.69	118.20
10	129.54	135.03	135.21
11	120.21	120.16	120.97
11a	132.74	137.10	137.52
12	187.53	187.84	190.71
12a	128.81	131.06	138.37 ^a
12b	140.03	140.08	119.94 ^b
13	29.11	29.41	21.19
14	56.04	56.54	56.62

Bruker AM 360, 90 MHz, CDCl₃, δ values (ppm) with CDCl₃ as an internal standard (δ 77.00).

^{a,b} Assignment exchangeable.

Table 2. ¹H NMR data of SM 196 A (1) and SM 196 B (2).

Proton	SM 196 A (1)	SM 196 B (2)
1-H	5.59 dddd (<i>J</i> =6.70, 5.87, 4.5, 0.5)	5.48 dddd (<i>J</i> =6.93, 6.07, 4.2, 0.5)
1-OH	5.20 d (<i>J</i> =4.5)	5.05 d (<i>J</i> =4.2)
2-H _a	2.16 ddd (<i>J</i> =-13.63, 5.87, 1.00)	2.15 (<i>J</i> =-13.86, 6.07, 1.00)
2-H _b	2.31 ddd (<i>J</i> =-13.63, 6.70, 1.50)	2.34 ddd (<i>J</i> =-13.86, 6.93, 1.65)
3-OH	1.48 (br)	2.0 (br)
4-H _a	2.90 dddq (<i>J</i> =-16.33, 1.50, 0.4)	2.91 dddq (<i>J</i> =-16.78, 1.65, 0.4)
4-H _b	3.09 ddddq (<i>J</i> =-16.33, 1.00, 0.5, 0.4)	3.10 ddddq (<i>J</i> =-16.78, 1.00, 0.5, 0.4)
5-H	7.48 dt (<i>J</i> =8.07, 0.4)	7.51 dt (<i>J</i> =8.04, 0.4)
6-H	7.85 d (<i>J</i> =8.07)	8.19 (<i>J</i> =8.04)
7-H	5.99 dd (<i>J</i> =2.0, 0.3)	
7-OH	4.13 d (<i>J</i> =2.0)	
9-H	7.20 ddq (<i>J</i> =8.15, 0.74, 0.2)	7.20 ddd (<i>J</i> =8.48, 0.96, 0.2)
10-H	7.50 ddd (<i>J</i> =8.15, 7.92, 0.3)	7.70 dd (<i>J</i> =8.48, 7.79)
11-H	7.92 dd (<i>J</i> =7.92, 0.74)	7.87 dd (<i>J</i> =7.79, 0.96)
13-H ₃	1.50 s (br)	1.50 s (br)
14-H ₃	4.04 d (<i>J</i> =0.2)	4.04 d (<i>J</i> =0.2)

Bruker AM 360, 360 MHz, CDCl₃, δ values (ppm) and *J* (Hz), TMS as internal standard, (¹H)=0.000 ppm, temperature: 303 K display value.

Table 3. Biological activities of 1 and 2 (MIC in $\mu\text{g/ml}$).

	<i>Staphylococcus aureus</i>	<i>Streptococcus pyogenes</i>		Herpes simplex 1	Herpes simplex 2
	H 503	S 308/7	S 77/7		
1	100.0	25.0	12.5	—	—
2	25.0	6.25	6.25	0.55	4.54

The dihydroquinone is rapidly oxidized by oxygen in the air to the corresponding yellow quinone SM 196 B (**2**). Simultaneous radiation (366 nm) led to the orange 6-deoxy-8-*O*-methylrabelomycin (**3**), which exhibits both a quinone moiety in ring B and a ketone moiety in ring D. The brilliant red color observed on the TLC plate after treatment with UV light derives from the dehydration product X-14881 E (**4**), which is to be distinguished from the other compounds by aromatization of ring D. Scheme 1 illustrates the isolation sequence of the SM 196 antibiotic complex. Due to its rapid conversion to **2**, **3**, and **4** on TLC plates it could be assumed that SM 196 A (**1**) is the native natural product. Therefore, no enzymatic steps seem to be involved in the dehydrogenation and dehydration reactions. To study the TLC-plate decomposition in more detail, these reactions were performed on preparative scale using more defined experimental conditions. Stirring of SM 196 A (**1**) in CHCl_3 in the presence of oxygen and in the absence of light the quinone **2** was formed within 4 hours. Performing the same reaction by irradiation of **1** with UV light (366 nm) for 6 hours yielded **3** in 60%. A side reaction to X-14881 E (**4**) was observed, which could be explained by hydrochloric acid impurities present in the solvent chloroform. This was proved by the conversion of **3** to **4** in chloroform/1% HCl in the dark.

The described angucyclinones SM 196 A (**1**) and (**2**) exhibit antibacterial⁹⁾ and antiviral activity¹⁰⁾ (see Table 3).

Addendum in Proof

Recently, an analog photo-induced oxidation as described for the antibiotics SM 196 A and B was reported for the rubiginones: OKA, M.; M. KONISHI, T. OKI and M. OHASHI: Absolute configuration of the rubiginones and photo-induced oxidation of the C-1 hydroxyl of the antibiotics to a ketone. *Tetrahedron Letters* 31: 7473~7474, 1990.

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